

# **Key considerations in balancing risk against need in Chinook supplementation**

**Kerry Naish,  
School of Aquatic and Fishery Sciences  
University of Washington**

# Recent Reviews of Hatchery Programs

## Responsible Approach to Marine Stock Enhancement: An Update

KAI LORENZEN,<sup>1,2,3</sup> KENNETH M. LEBER,<sup>3</sup> and H. LEE BLANKENSHIP<sup>4</sup>

### Ecological interactions between wild and hatchery salmonids: an introduction to the special issue

Peter S. Rand • Barry A. Berejikian •  
Todd N. Parsons • David L. G. Noakes

## Factors that contribute to the ecological risks of salmon and steelhead hatchery programs and some mitigating strategies

Kathryn Kostow

## Hatcheries, Conservation, and Sustainable Fisheries—Achieving Multiple Goals: Results of the Hatchery Scientific Review Group's Columbia River Basin Review

P. J. Paquet<sup>a</sup>, T. Flagg<sup>b</sup>, A. Appleby<sup>c</sup>, J. Barr<sup>d</sup>, L. Blankenship<sup>e</sup>, D. Campton<sup>f</sup>, M. Delarm<sup>g</sup>, T. Evelyn<sup>h</sup>, D. Fast<sup>i</sup>, J. Gislason<sup>j</sup>, P. Kline<sup>k</sup>, D. Maynard<sup>l</sup>, L. Mobrand<sup>m</sup>, G. Nandor<sup>n</sup>, P. Seidel<sup>o</sup> & S. Smith<sup>p</sup>

# Main conclusions from review



## **AN EVALUATION OF THE EFFECTS OF CONSERVATION AND FISHERY ENHANCEMENT HATCHERIES ON WILD POPULATIONS OF SALMON<sup>1</sup>**

Kerry A. Naish,<sup>\*2</sup> Joseph E. Taylor, III<sup>†</sup> Phillip S. Levin,<sup>‡</sup> Thomas P. Quinn,<sup>\*</sup> James R. Winton,<sup>§</sup> Daniel Huppert,<sup>¶</sup> and Ray Hilborn<sup>\*</sup>

*Advances in Marine Biology*, Volume 53  
ISSN 0065-2881, DOI: 10.1016/S0065-2881(07)53002-6

© 2008 Elsevier Ltd.  
All rights reserved.

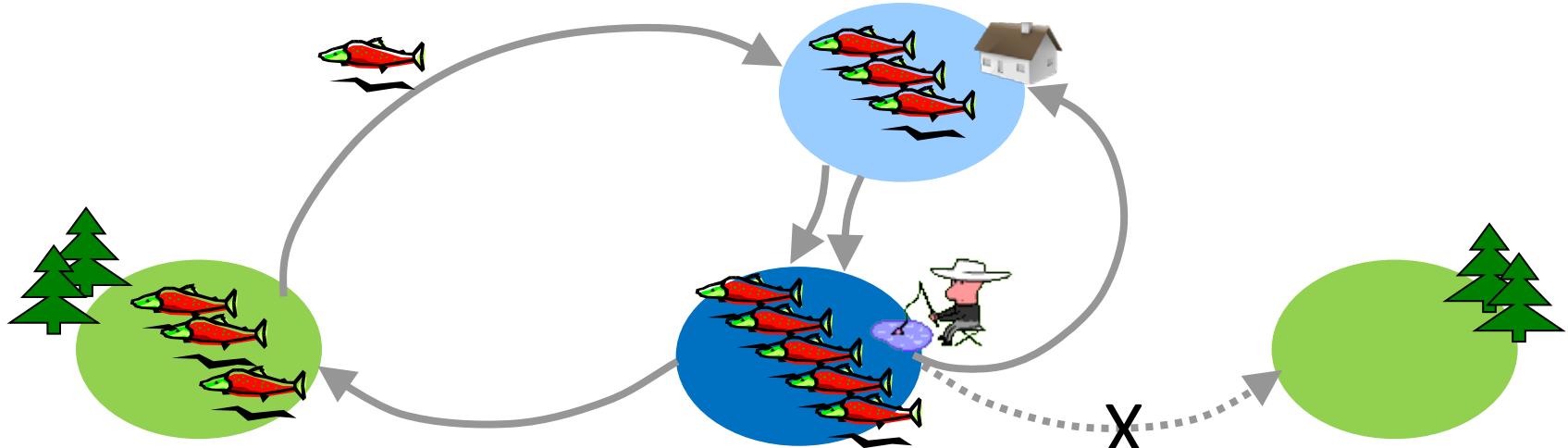
- “A key need in this area..... is the development of a strong understanding of the *degree* to which specific activities pose a risk and whether proposed management approaches are effective at reducing these risks”

# Aims

1. Key discussion associated with minimizing risks
2. Examples of outstanding issues
3. Considerations for planning a new program
  - Towards a well designed system that allows research and continual assessment

# Release objectives

- Produce surplus fish for fishing opportunity



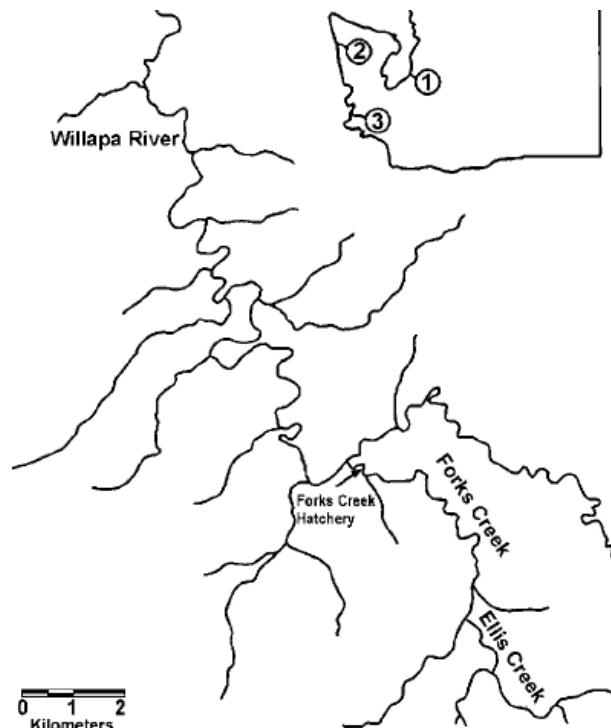
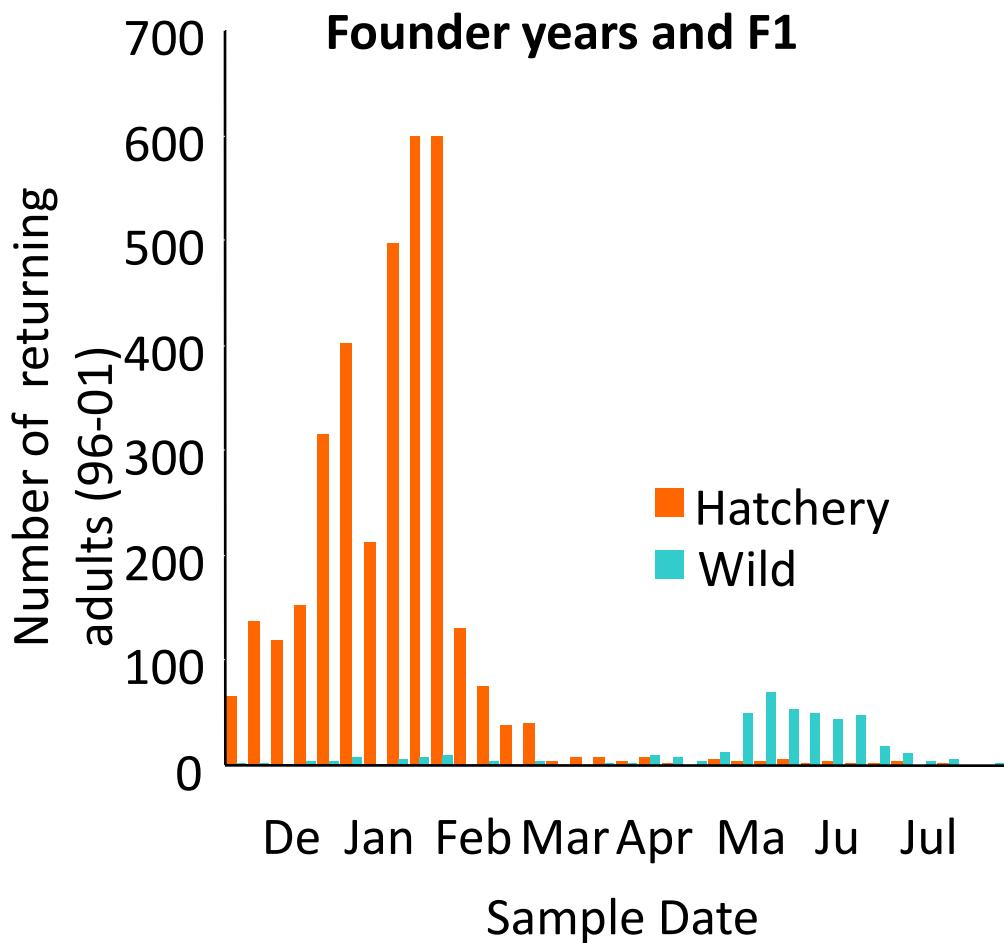
## ***Integrated Hatchery Aims:***

- Genetic constitution and fitness as similar as possible to wild population
- Achieved by gene flow:
  - Wild to hatchery > hatchery to wild

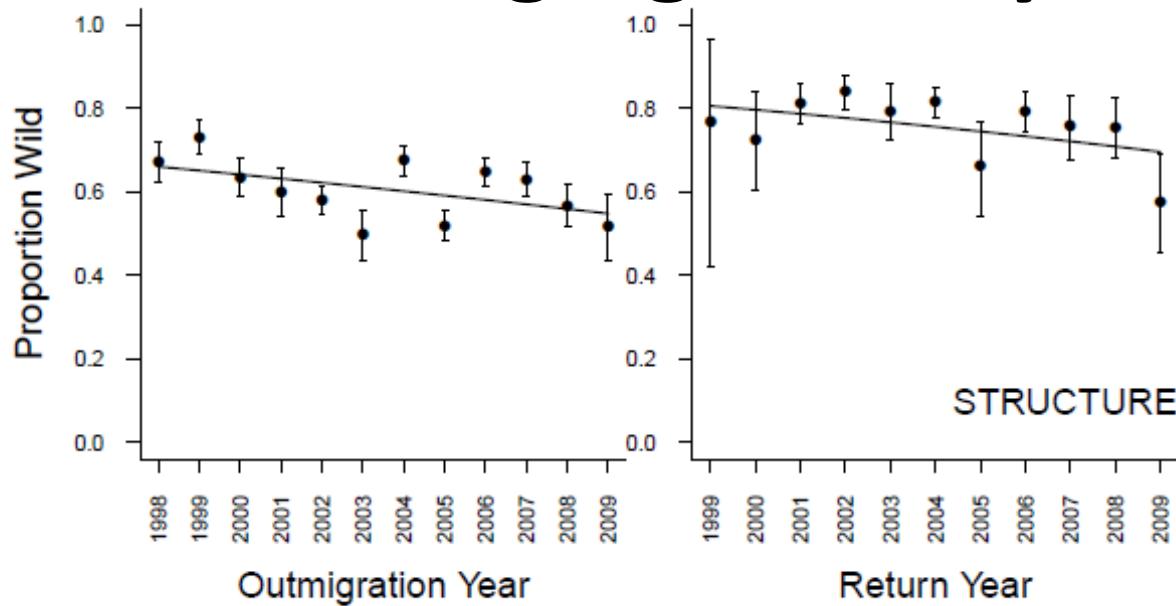
## ***Segregated Hatchery Aims:***

- Direct fishing effort away from wild populations
- Little or no spawning in the wild

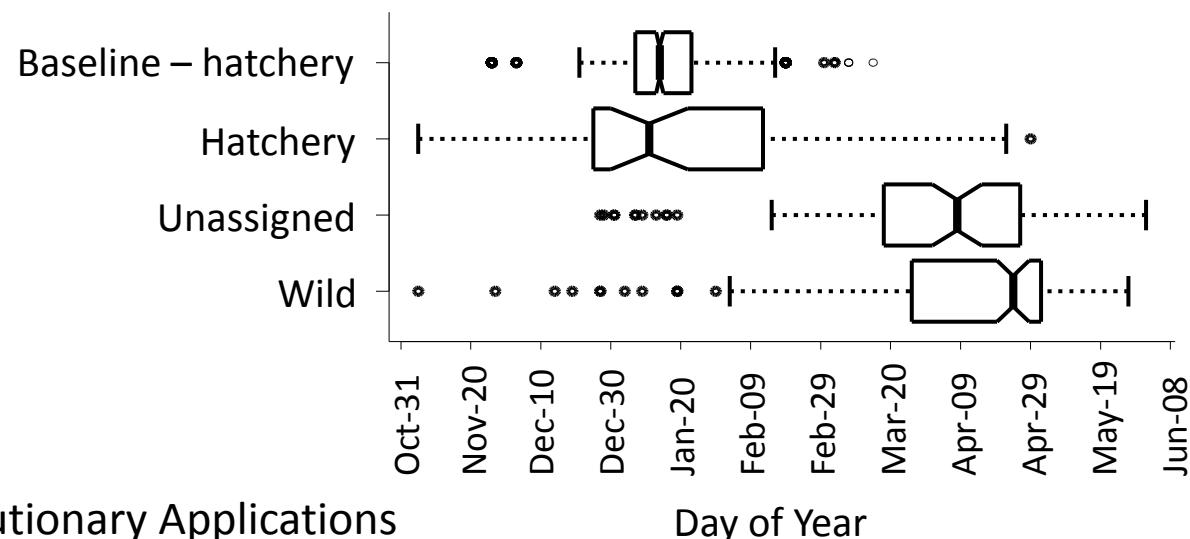
# Segregation – a viable approach?



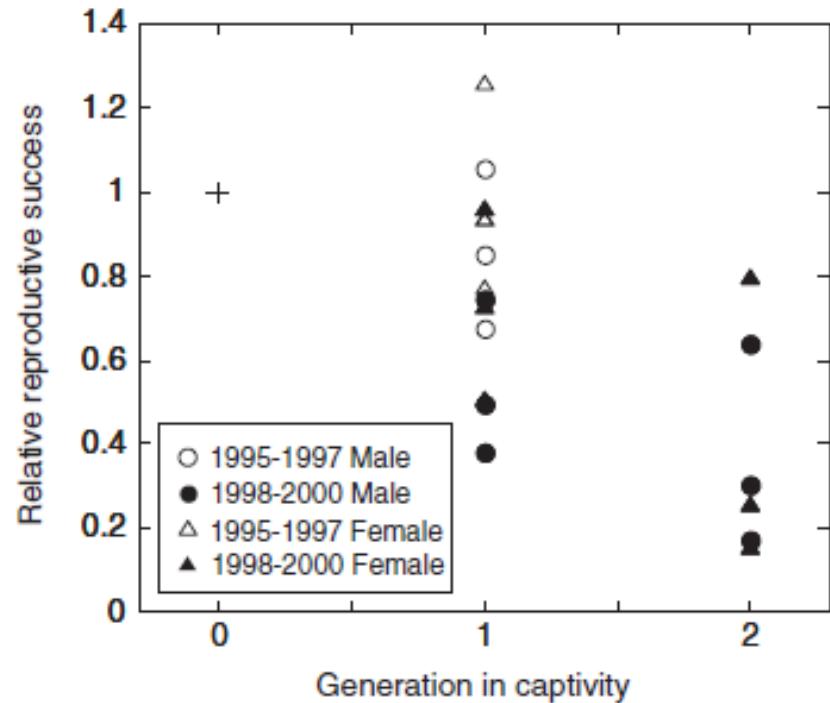
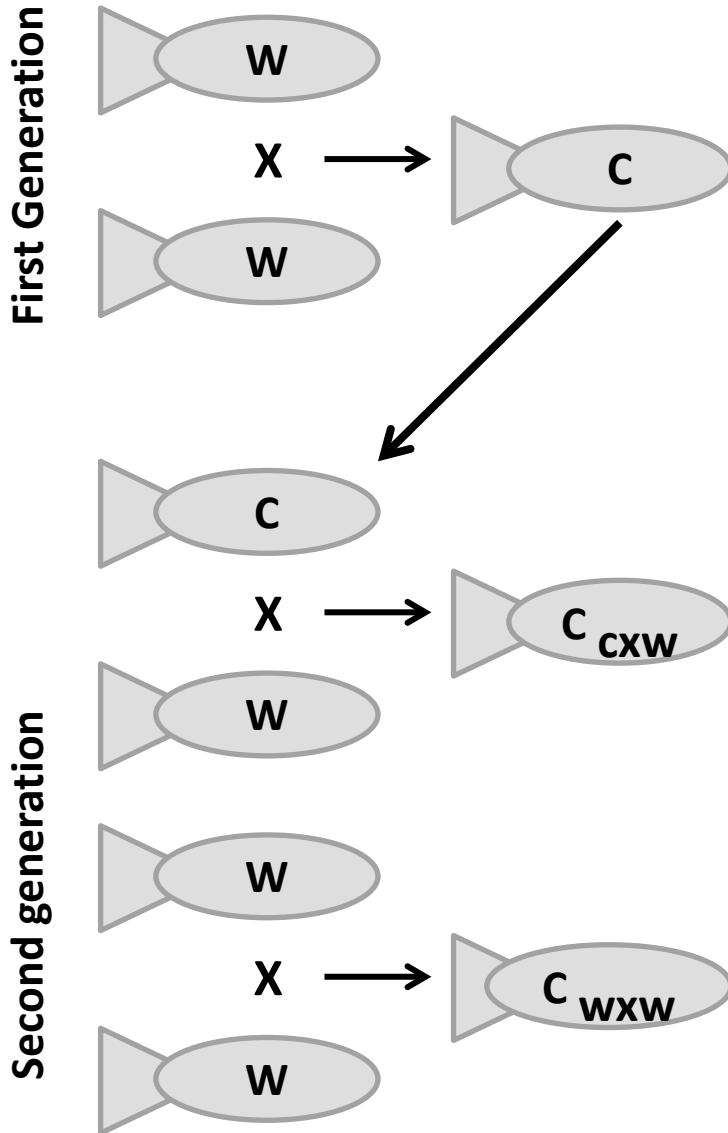
# Does segregation by timing work?



STRUCTURE



# Integrated hatcheries: number generations?



- Decline in fitness observed in a steelhead hatchery over two generations following captive rearing
- Araki, Cooper & Blouin, *Science* 2007

# Integrated: subsequent studies

Species	Outcome	References
Steelhead	<ul style="list-style-type: none"><li>Reduced fitness in H descendants spawning in W reduce d W pop size,</li><li>Swamping: 40% contributions from W</li><li>Domestication selection in one generation</li></ul>	Araki et al. 2008, Christie et al. 2011, Christie et al. 2012
	<ul style="list-style-type: none"><li>No changes in genetic diversity or effective size (Adult to parr)</li></ul>	Van Doornick et al. 2010
	<ul style="list-style-type: none"><li>Methodology: RRS estimates are upwardly biased</li></ul>	Kitada et al. 2011
Chum	H & W similar fitness after 3 generations of culture (spawning channel)	Berejikian et al. 2009

# Integrated: subsequent studies

Species	Outcome	Reference
Coho	<ul style="list-style-type: none"><li>Rearing environment (natural vs trad) affects fitness</li><li>No genetic explanation for fitness decrease</li><li>H fish had lower RRS</li><li>Fitness of fry release similar to smolt release</li><li>Absence of sexual selection in H implicated</li></ul>	<p>Chittenden et al. 2010</p> <p>Theriault et al. 2011</p>
Chinook	<ul style="list-style-type: none"><li>No fitness difference after one generation of H rearing (lab)</li><li>H fitness lower than W in established supplementation hatchery</li><li>H fish younger, spawned in different habitat</li><li>H fish over a single generation provided a demographic boost</li><li>No significant difference in RRS in H&amp;W spawning in W</li></ul>	<p>Schroder et al. 2010, 2012</p> <p>Williamson et al. 2011</p> <p>Hess et al. 2012</p>

# Aim of Talk

1. Key discussion associated with minimizing risks
2. Examples of outstanding issues
3. Considerations for planning a new program
  - Towards a well designed system that allows research and continual assessment

# Integrated: outstanding questions

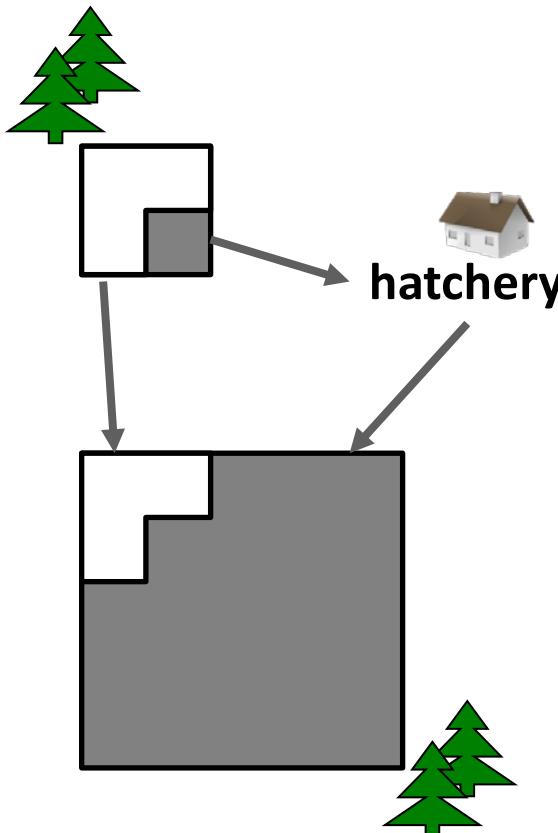
- What are measures of success in supplementation?
  - Is reduced fitness in descendants of hatchery fish acceptable or even desirable?
- Optimal levels of gene flow between Hatchery and Wild fish? Need for experimental approach
- Effects of broodstock collection on wild fish?
- Does long term integration eventually compromise the genetic diversity of a wild population?
- Are hatcheries at risk of becoming pathogen vectors?

# Segregated: outstanding questions

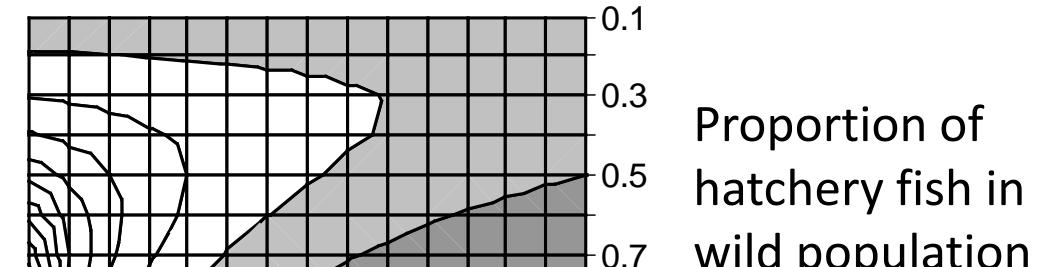
- Are there design strategies that prevent negative interactions between hatchery and wild fishes?
- How much gene flow is acceptable if systems fail?
- Effects of broodstock collection on wild fish?
- Mixed stock fisheries
- Are hatcheries at risk of becoming pathogen vectors?

# The importance of release sizes

- May affect ability of wild populations to adapt to change
  - Reduction of genetic diversity: the “Ryman – Laikre” effect



**Effective size of hatchery fish = 100**

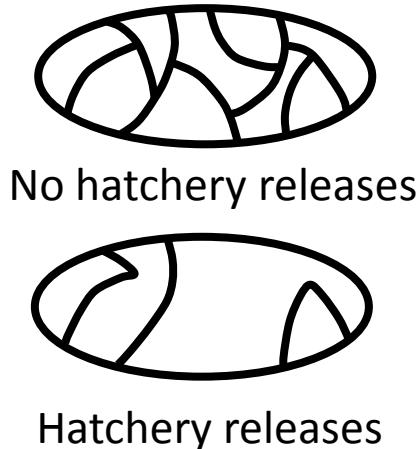


Effective size of wild population

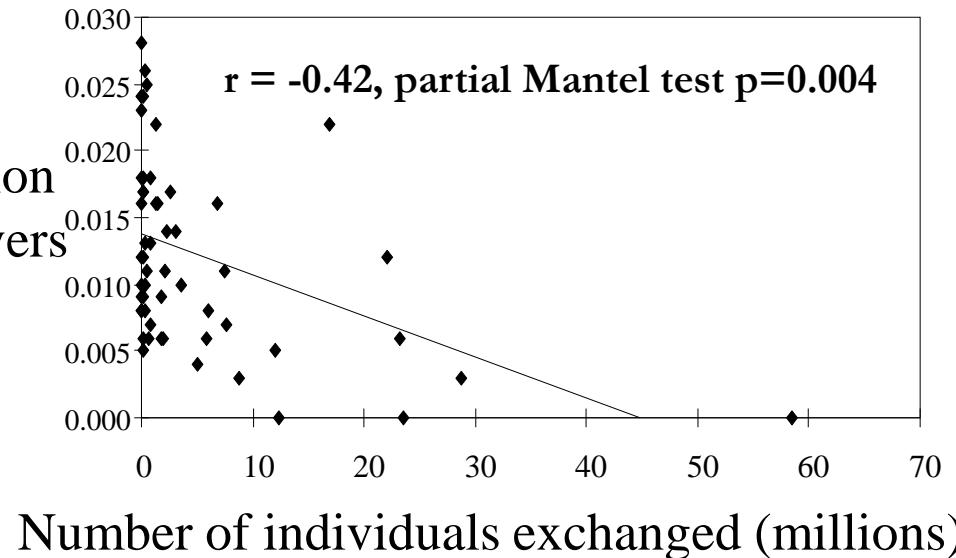
Grey = reduction in effective size  
of total population compared to  
unsupplemented population

# The importance of release sizes

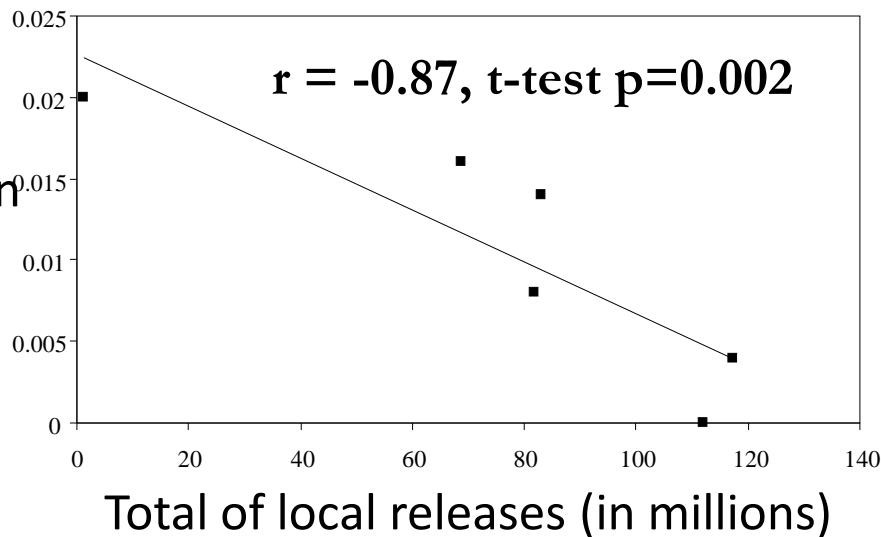
- May reduce population structure (*Utter, 2004*)



$F_{RT}$ ,  
differentiation  
between Rivers



$F_{SR}$  Genetic  
differentiation  
within rivers



*Eldridge and Naish 2007, Molecular Ecology*  
*See also Marie et al. 2010, Molecular Ecology*

# Release sizes: Ecological effects

“Many hatchery management strategies that may decrease genetic risks, ...., (but they) may not mitigate ecological risk factors, rather they may increase the opportunity for ecological effects to occur.” *Kostow 2010*

- Competition, exacerbated by physical differences
- Density dependent mortality
- Residualization
- Carrying capacity, especially in freshwater
- Predation
- Series of recommendations to reduce ecological effects

# Release sizes: Ecological effects

“the traditional hatchery paradigm is to release approximately the same number of fish of the same species every year from the same location(s) regardless of ecological conditions. This paradigm occurs in integrated and segregated hatchery programs and generally ignores ecological feedback mechanisms within the environment that can reduce survival of hatchery and wild fish” *Pearsons 2010*

Release strategies should consider;

- Risks to non-target taxa
- Carrying capacity
- Ecological feedback mechanisms

# Aim of Talk

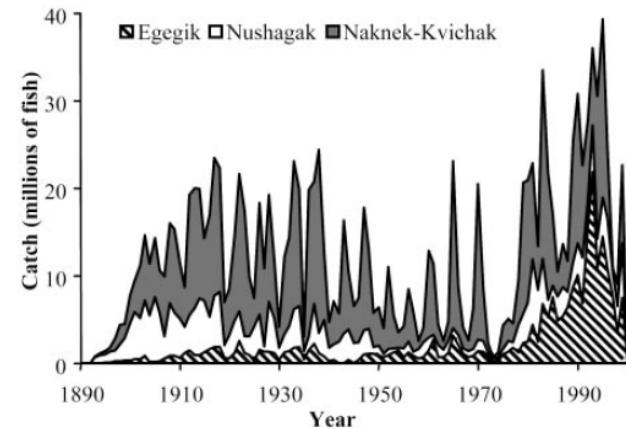
1. Key discussion associated with minimizing risks
2. Examples of outstanding issues
3. Considerations for planning a new program
  - Towards a well designed system that allows research and continual assessment

# Planning a hatchery program

- Stage I: Initial appraisal and goal setting
  - Release objectives
  - Interactions with wild fish
  - Importance of release sizes
- Stage II: Research and technology development including pilot studies
  - Generating population baselines
  - Designing an experimental system
  - Tagging and tracking hatchery and wild fish
- Stage III: Operational implementation and adaptive management

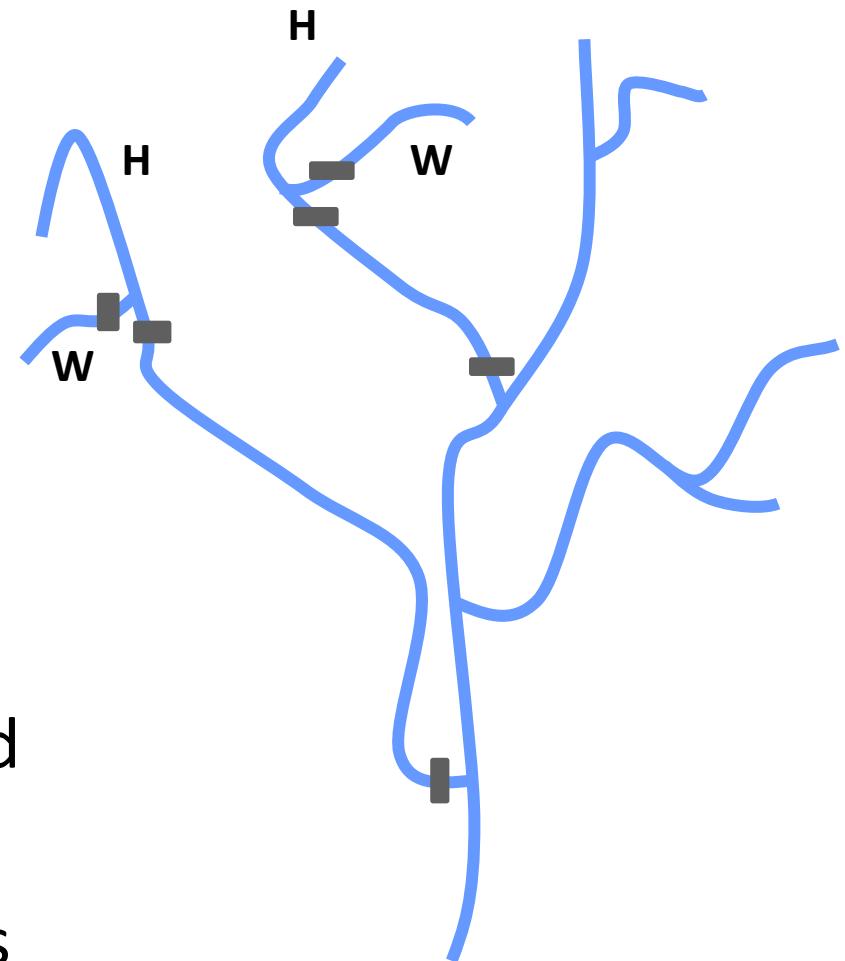
# Knowledge gaps: Baseline data sets

- Maintaining the “portfolio”
  - (*Hilborn et al 2003, Schindler et al. 2010*)
  - Evidence for population structure?
  - Evidence for local adaptation?
  - Evidence for life history variation within and between populations?
  - Estimate migration rates between populations
  - Evidence for changes in structure over time



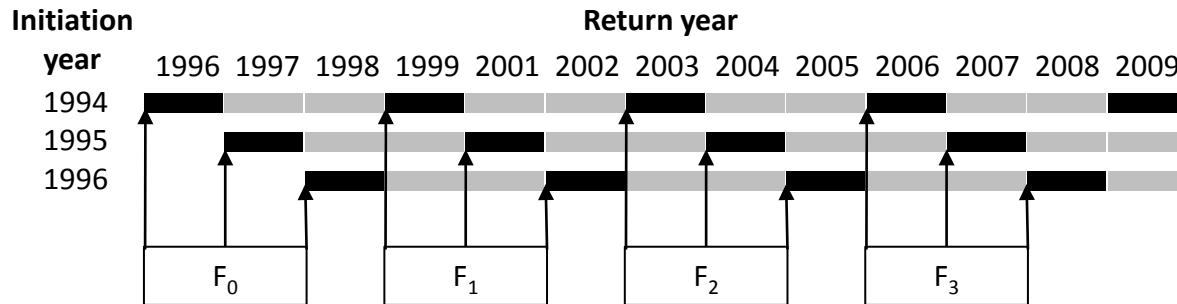
# Knowledge gaps: feasibility study

- System that permits effective ongoing research
  - Control streams, free of hatchery influence
  - Continual evaluation
    - “Trigger points”
    - Decide how much change is acceptable
  - Willingness to change or end practices
  - Economic and social analysis

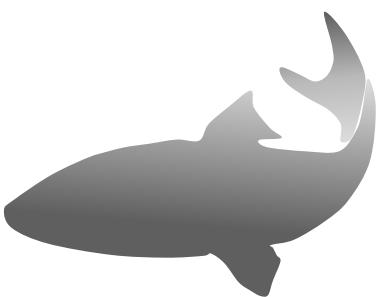


# The role of comprehensive long term data

- Measuring phenotypes and pedigrees

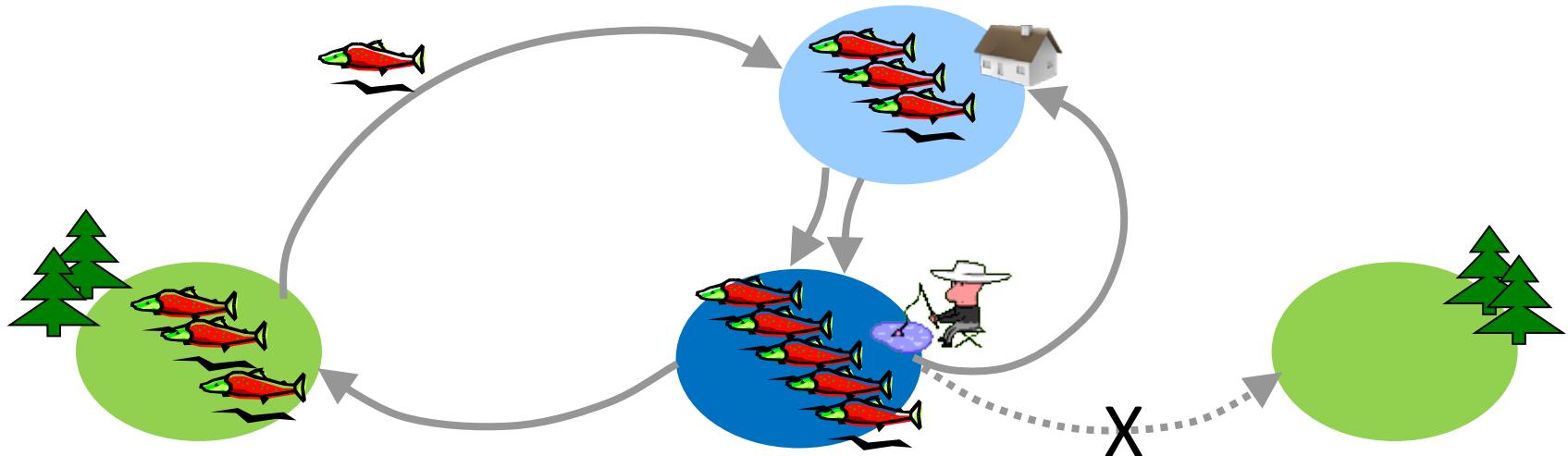


- Pedigree data - beyond RRS :
  - Determine hatchery ancestry and contribution over time (eg Christie et al. 2011)
  - Effective population sizes, rates and impacts of inbreeding (eg Naish et al. Molecular Ecology in press)
  - Estimate selection differentials in hatchery and wild environments (eg Ford et al. 2010, Williamson et al 2011)
  - Estimate stray rates between populations (eg Lin et al. 2011, in prep.)



**Thank you!**

# Summary: balancing risks



## ***Integrated Hatchery Risks:***

- Proportion of Hatchery fish breeding in the wild
- Optimal number of generations in program
- Optimal number of wild fish in program (broodstock mining)
- Carrying capacity, competition

## ***Segregated Hatchery Risks:***

- Divergent hatchery fish interbreed with wild
- Ecological interactions, carrying capacity
- Mixed stock fisheries and efficacy of mass marking

# Summary: Considerations in design

- A research based approach
  - Test principles as well as specifics
  - Continual evaluation
  - “Trigger” points for changing or ending practices
- Comprehensive baselines and monitoring program