Alaska Hatchery Research Program

- Why the program was initiated
- Program structure
- Key questions addressed
- Study design

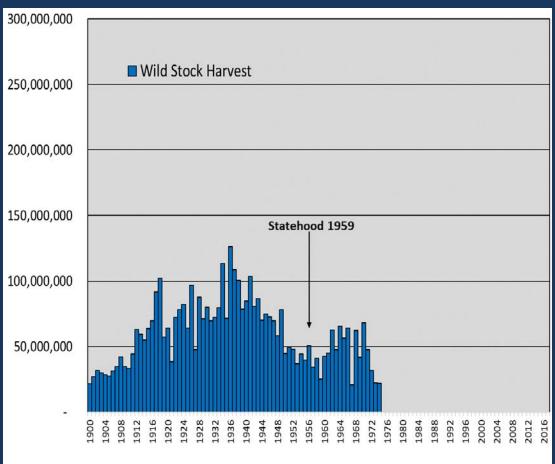


Background/Salmon Enhancement Issues



Background

 Alaska salmon fisheries were severely depressed at statehood, and reached their nadir in 1973 and 1974, when statewide harvest of all species was 22 M

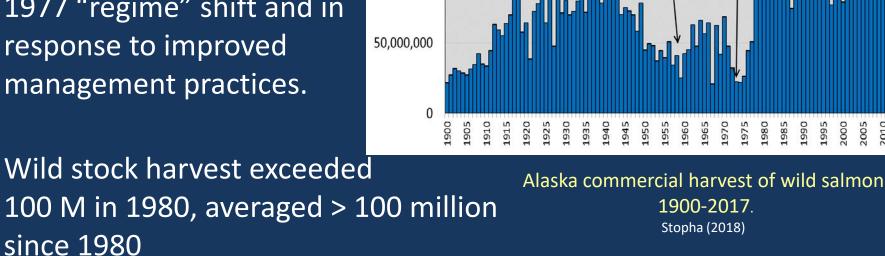


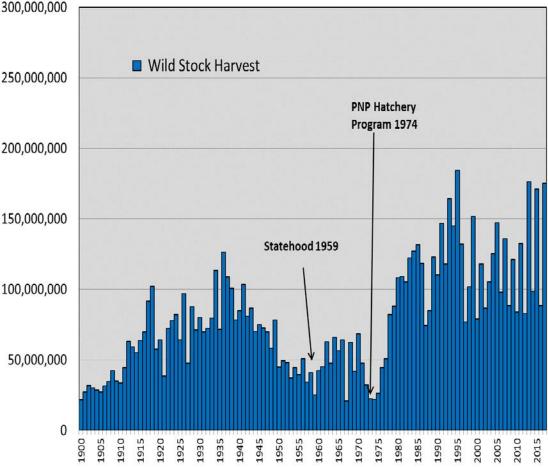
Alaska commercial salmon harvest 1900-1974 Stopha (2018)

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Background

- Alaska initiated State (1971) 300,000,000 and PNP (1974) hatchery programs to support the recovery and enhancement of Alaska salmon fisheries.
- Remarkable renaissance of Alaska salmon following 1977 "regime" shift and in response to improved management practices.



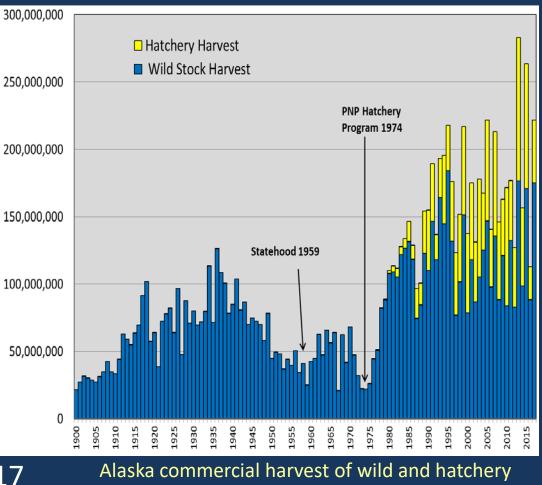


Background

 Hatcheries began making substantial contributions to harvest in 1980's

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- Statewide harvests (wild and hatchery) have averaged 175 M annually for 2008-2017
- Hatcheries produced an annual average of 67 M, 33% of the harvest, 2008-2017



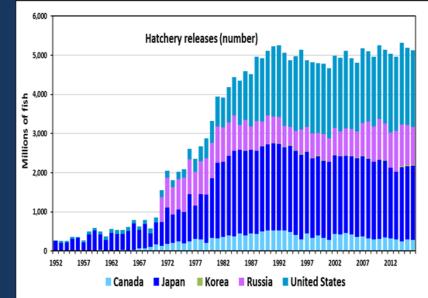
salmon, 1900-2017. Stopha (2018)

 Hatchery production now dominates the harvest of pink and chum salmon in PWS and chum salmon in SEAK

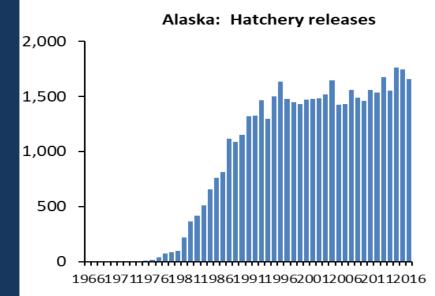
BACKGROUND

Large-scale salmon releases raise concerns for wild stock impacts

- Do hatchery fish detrimentally affect productivity and sustainability of wild stocks?
- Alaska policy mandates sustainable productivity of wild stocks
- Not a new concern: Alaska first state to have a Genetics Policy in 1985



Data Source: North Pacific Anadromous Fish Commission (NPAFC). 2017. NPAFC Pacific salmonid hatchery release statistics (updated 31 July 2017). North Pacific Anadromous Fish Commission, Vancouver. Accessed July, 2017. Available: www.npafc.org



ADF&G Genetic Policy

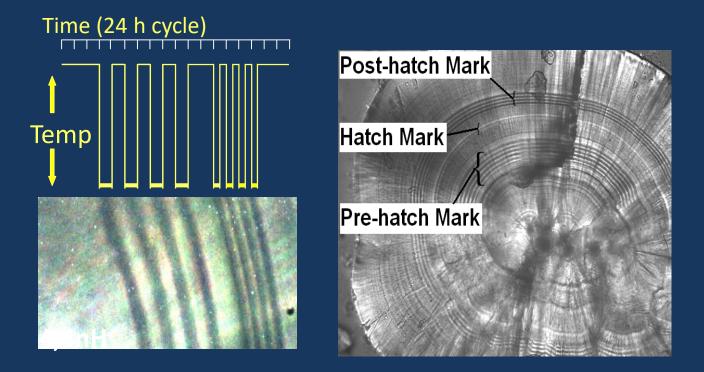
Contains protections for wild stocks while allowing increased productivity for enhancement programs.

Image: marginal states in the states of the t

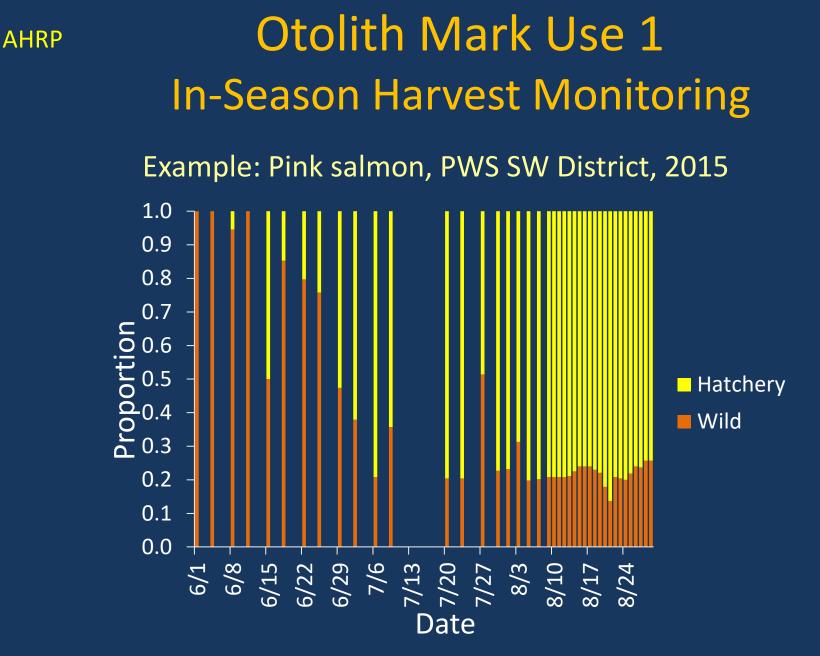
- Reduce gene flow from hatchery to wild
- Minimize introgression of ill-adapted genes
- Minimize hybrid depression
- Maintain stock fitness
- Minimize magnitude of straying
- Temporal and spatial isolation are important



AHRP Tool for identifying hatchery fish Otolith Thermal Marking



Alaska marks > 80% of hatchery fish ~ 1.2 Billion (100% for PWS and SEAK pink and chum)



Haught, S., J. Botz, S. Moffitt, and B. Lewis. 2017. 2015 Prince William Sound area finfish management report. Alaska Department of Fish and Game, Fishery Management Report No. 17-17, Anchorage.

Otolith Mark Use 2 Measure Straying

- Are hatchery fish straying? If so, how many fish are straying?
- SE Alaska chum Heinl and Piston (2008-2010)
- Prince William Sound pink, chum & sockeye

Joyce and Evans (1997-1999) Brenner and Moffitt (2004-2010)

 These studies found widespread distribution of hatchery strays in their respective regions, and high rates of hatchery strays in streams near hatchery release sites



AHRP Recognition of need to examine extent and impact of hatchery strays on wild stock fitness and productivity

- PNP operators proposed that ADF&G organize a science panel of experts to design and implement a long term research project to inform future resource management decisions
- Funding partnership: State, Operators & Industry
- Fundamental questions aimed at examining extent and potential impacts of hatchery straying on fitness of wild stocks
 - * Pink and chum salmon PWS
 - * Chum salmon SEAK

AHRP Science Panel

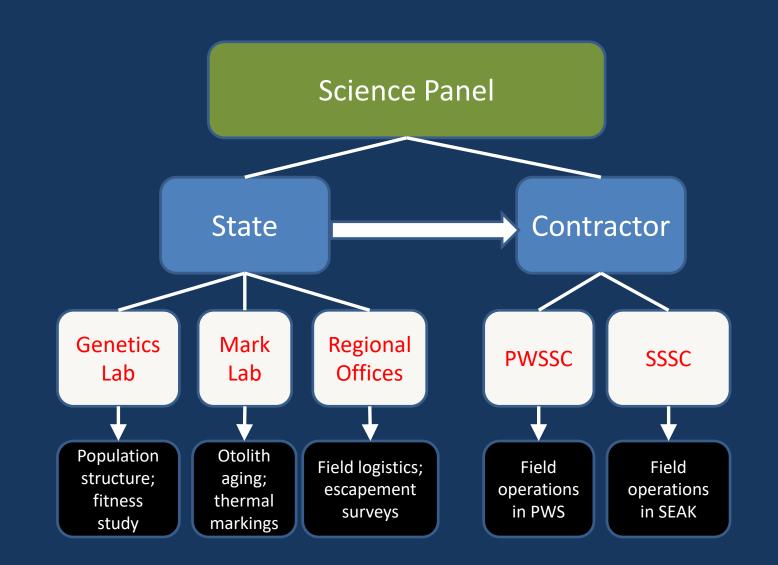
Panel Charge –

Identify priority research questions and develop a framework for research that could be used to address these questions.

Panel Makeup – 13 members:

- Alaska Department of Fish and Game
- National Marine Fisheries Service
- University of Alaska
- Aquaculture associations

AHRP Structure





- 1) What is the genetic stock structure of pink and chum in PWS and SEAK?
- 2) What is the extent and annual variability of straying?
- 3) What is the impact on fitness (*productivity*) of natural pink and chum stocks?



- 1) What is the genetic stock structure of pink and chum in PWS and SEAK?
- Hatchery broodstocks are all derived from stocks within their respective regions as per the Alaska Genetics Policy
- Broodstocks are necessarily large because of the scale of the program, and brood fish are spawned non-selectively from throughout the duration of the spawning run.
- As a result, broad-scale genetic variability is maintained, but the distribution of genotypes may differ from wild spawning populations which are smaller and potentially adapted to localized environmental conditions.
- Policy makers and resource managers need an improved understanding of the hierarchical genetic structure of the populations, and the degree of genetic differentiation at and within the level of harvest management resolution.

AHRP Research Questions

1) What is the genetic stock structure of pink and chum in PWS and SEAK?

Pink salmon in PWS

- Ecologically important, but shallow, structure observed in even and odd years (1990s)
- Re-examine structure with new samples and new markers (2013-2015)
- Compare 1990s structure to present structure

Chum salmon in PWS and SEAK

- Temporal and regional structuring observed within SEAK and PWS (1990s & 2013).
- Collect additional samples to supplement baseline collections
- Examine fine-scale structure using updated methods



1) What is the genetic stock structure of pink and chum in PWS and SEAK?

- Research directed by the ADF&G Gene Conservation Laboratory
- Collaborative effort with University of Alaska, NOAA Auke Bay Laboratories, University of Washington, Prince William Sound Science Center and Sitka Sound Science center
- Results to provide perspective on degree of heterogeneity across different geographic scales, insight into temporal changes associated with hatchery production, and improved capacity to track future changes.

2) What is the extent and annual variability of Hatchery straying?

- Stray Rate (1): Proportion of a spawning population that spawns in a stream other than its natal stream
- Stray Rate (2): Proportion of a stream's escapement that is composed of strays from other streams (or hatcheries)
- Estimation of hatchery stray rate (2): Proportion of spawning composed of hatchery-origin fish at the stream, management district, and region level for pink salmon and chum salmon in PWS and chum salmon in SEAK

2) What is the extent and annual variability of Hatchery straying?

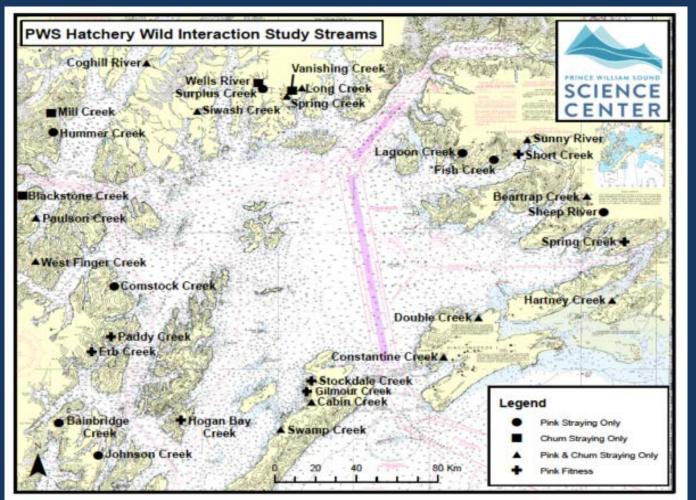
- Randomly select streams with selection weighted for escapement within and between management districts so that stray rates in streams can be used to accurately estimate district and regional rates
- Initial target of 384 fish per stream to estimate stray rate within 5% with 95% confidence
- Sample otoliths from carcasses throughout run, weighting samples in calculating hatchery proportion by the carcass counts at each sampling event

2) What is the extent and annual variability of hatchery salmon straying?

PWS Pinks:
 27 streams
 8 districts

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 PWS Chums: 17 streams
 6 districts



2) What is the extent and annual variability of hatchery salmon straying?

SEAK Chums:
 32 streams
 3 sub-regions

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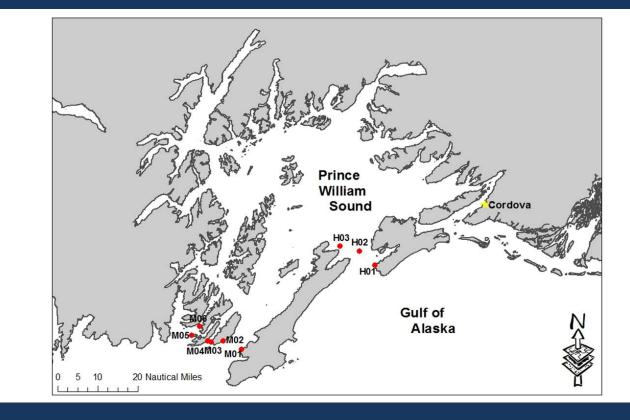
2) What is the extent and annual variability of hatchery salmon straying?

- In addition to estimation of proportion of hatchery strays in spawning escapement, the study was designed to estimate:
 Number of wild spawners
 Number of hatchery strays
 - -Production (total run) of wild and hatchery fish
- Can be calculated from catch numbers (known), hatchery broodstock numbers (known), proportion hatchery fish in spawning escapement (estimated), and PROPORTION OF HATCHERY FISH IN TOTAL RUN
- Estimate in PWS with ocean sampling as fish enter Sound
- Estimate in SEAK from chum salmon incidental catch in pink salmon seine fishery: not feasible due to data resolution in fishery

2) What is the extent and annual variability of hatchery salmon straying?

Ocean sampling 2013–2015 (PWS only)

- Fish entering PWS sampled at 9 stations
- Multi-mesh panel gillnet fished from contracted gillnet vessel
- Twice weekly sampling from mid-May until the end of August

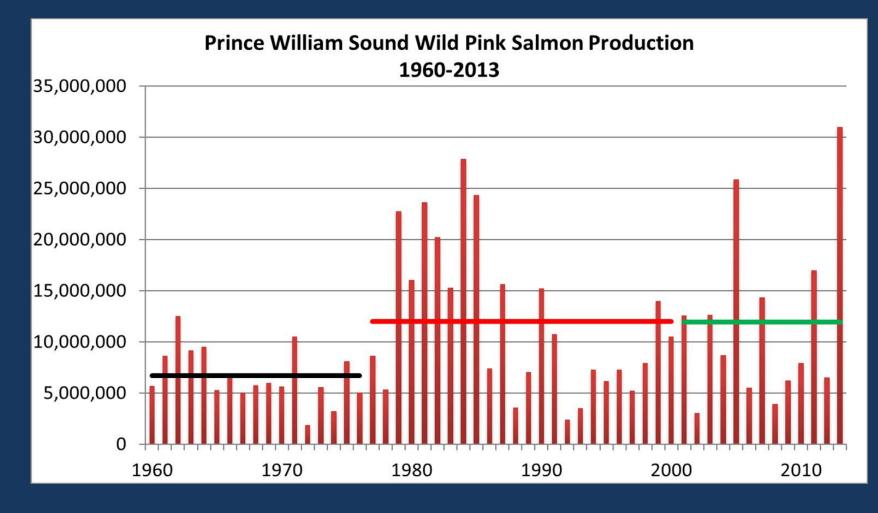


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2) What is the extent and annual variability of hatchery salmon straying?

- Ocean sampling allows estimates of total hatchery and wild runs, wild escapement, and number of hatchery strays in the escapement.
- Can compute the hatchery stray rate (1): proportion of hatchery run that strays into spawning streams
- Can contrast production with estimates using index counts to estimate escapement

2) What is the extent and annual variability of straying?



From Gaudet (2017)

3) What is the impact on fitness (productivity) of natural pink and chum stocks?

Fitness – the ability to survive and reproduce [average contribution by average individual to next generation]

If hatchery fish are less fit in wild streams, then

1. Hatchery fish will produce fewer offspring

AND

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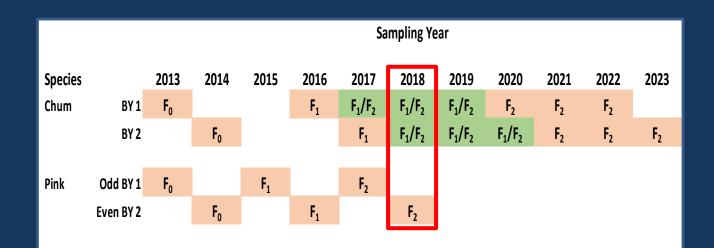
2. Wild fish will produce fewer offspring due to interbreeding.

3) What is the impact on fitness (productivity) of wild pink and chum stocks?

- Identify number of offspring produced by hatchery- and wild-origin parents
- Follow 2 brood years for 2 generations: replicate for F₁, potential for F₂
- PWS pink salmon: 6 streams (2013-2018)

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- SEAK chum salmon: 4 streams (2013-2023)
- Target streams with high (@50%) and low (@10-20%) hatchery stray rates



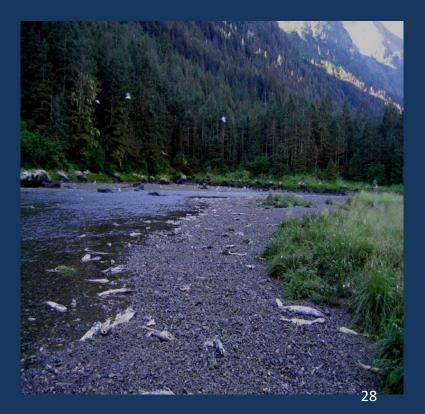
Otolith Mark Use 3

Are there effects of straying on fitness and productivity?

- Use otolith marks to identify hatchery vs. natural origin spawners
- Use parentage analysis to assess fitness of hatchery and wild fish
- Studies in Pacific NW on other species may not apply to AK (king, coho, steelhead)
 - ✓ Freshwater residence time
 ✓ Life span and age structure
- Different hatchery practices

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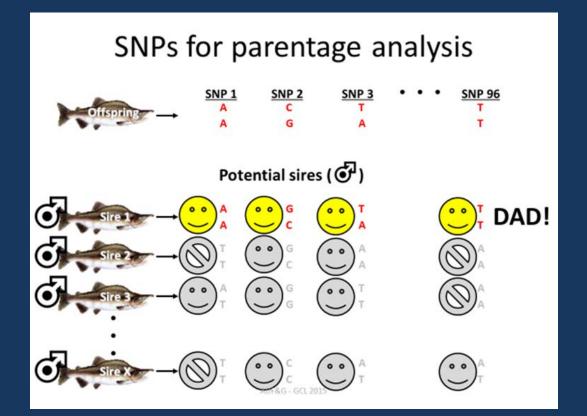
- ✓ Local broodstock in AK
- ✓ 10,000 + parents spawned
- Limited holding or feeding for pinks and chums



3) What is the impact on fitness (productivity) of natural pink and chum stocks? Parentage analysis – use genotyping of parents (F_0) for pedigree reconstruction to identify offspring in F_1 (and

F₂ if feasible) using single nucleotide polymorphisms (SNPs)

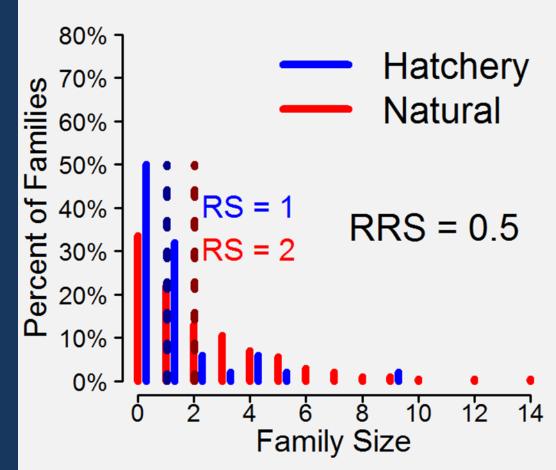
- SNPs available for chum salmon genotyping
- Extensive SNP development required for pink salmon genotyping
- Identify reproductive success (RS) of hatchery and wild parents



3) What is the impact on fitness (productivity) of natural pink and chum stocks?

Hypothetical distribution of family size for hatchery and wild parents

- Determine RS for hatchery and wild parents for each gender in a stream.
- Calculate the ratio (RRS)= RS(h)/RS(w)
- Differences in fitness must be large in order to detect as statistically significant; use RRS=0.5 for power analysis



3) What is the impact on fitness (productivity) of wild pink and chum stocks?

Power increases with...

• In our control

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- \uparrow Number families
- Stray rate > 10%
- \uparrow Proportion offspring
- Out of our control
 - Distribution of RS
 - 个 Mean
 - ↑ Dispersion
 - $-\downarrow$ True RRS

Depends on:

- Number parents (F₀) sampled
 Hatchery ~ f(stray)
 - Natural
- Proportion offspring (F₁) sampled
- Distribution of RS (productivity)
 - Mean
 - Dispersion
- RRS
 - Difference between Hatchery and Wild
 - Benchmark RRS = 0.5

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3) What is the impact on fitness (productivity) of wild pink and chum stocks?

Alevin Sampling

- Sampling alevins (post-hatch/pre-emergent larval fish) can provide insight into life-history stage of effects: freshwater versus marine
- Hydraulic pumping in spring to collect alevins that could be progeny of parents sampled after spawning the previous fall
- 250 sample sites distributed in relation to observed spawning abundance, with up to 25 aelvins retained per sample site
- Limit alevin sampling to one pink stream (Stockdale) and one chum stream (Fish Creek) due to budget constraints

AHRP AHRP AHRP Research Questions 3) What is the impact on fitness (productivity) of wild pink and chum stocks? Who does the work?

- Sampling of fitness streams contracted to PWSSC for PWS pink salmon, SSSC for SEAK chum salmon
- Otolith samples processed by ADF&G in Cordova (PWS) and Juneau (SEAK)
- SNP development for pink salmon a collaboration with University of Washington and ADF&G Gene Conservation Lab
- Processing and analysis of parentage and offspring samples the responsibility of the ADF&G Gene Conservation Lab